

```
1 from google.colab import drive  
2 drive.mount('/content/drive')  
3  
→ Drive already mounted at /content/drive; to attempt to forcibly remount, call drive.mount("/content/drive", force_remo
```



1. Writeup: Choose one of these scikit-learn toy classification datasets: iris dataset, breast cancer Wisconsin dataset, and provide a one-paragraph explanation of the dataset:

https://scikit-learn.org/stable/datasets/toy_dataset.html

Breast Cancer Wisconsin dataset

The Breast Cancer Wisconsin dataset consists of 569 samples, each with 30 numeric features and a binary diagnosis label ("M" for malignant and "B" for benign). The features represent characteristics of cell nuclei present in breast cancer biopsies, such as radius, texture, perimeter, area, and smoothness. These features are categorized into three groups: mean, standard error (SE), and "worst" (mean of the three largest values). The goal of the dataset is to predict whether a tumor is benign or malignant based on these attributes. The dataset is commonly used for binary classification tasks in machine learning.

```
1 zip_file_path = '/content/drive/My Drive/Datasets/breast+cancer+wisconsin+diagnostic.zip'  
2
```

```
1 with zipfile.ZipFile('/content/breast+cancer+wisconsin+diagnostic.zip', 'r') as zip_ref:  
2     zip_ref.extractall(extract_dir)  
3
```

```
1 import zipfile  
2 import os  
3
```

```
4 extract_dir = '/content/extracted_files/'  
5  
6 with zipfile.ZipFile('/content/breast+cancer+wisconsin+diagnostic.zip', 'r') as zip_ref:  
7     zip_ref.extractall(extract_dir)  
8  
9 extracted_files = os.listdir(extract_dir)  
10 print(extracted_files)  
11
```

```
→ ['wdbc.data', 'wdbc.names']
```

Double-click (or enter) to edit

2. Coding: Fetch the dataset as a dataframe

```

1 import pandas as pd
2
3 # Define the path to the extracted data file
4 data_file_path = '/content/extracted_files/wdbc.data'
5
6 # Load the dataset
7 column_names = [
8     "ID", "Diagnosis", "Radius_mean", "Texture_mean", "Perimeter_mean", "Area_mean", "Smoothness_mean",
9     "Compactness_mean", "Concavity_mean", "Concave_points_mean", "Symmetry_mean", "Fractal_dimension_mean",
10    "Radius_se", "Texture_se", "Perimeter_se", "Area_se", "Smoothness_se", "Compactness_se", "Concavity_se",
11    "Concave_points_se", "Symmetry_se", "Fractal_dimension_se", "Radius_worst", "Texture_worst", "Perimeter_worst",
12    "Area_worst", "Smoothness_worst", "Compactness_worst", "Concavity_worst", "Concave_points_worst",
13    "Symmetry_worst", "Fractal_dimension_worst"
14 ]
15
16 # Load the data with specified column names
17 df = pd.read_csv(data_file_path, header=None, names=column_names)
18
19 # Display the first few rows of the dataset
20 df.head()
21

```

	ID	Diagnosis	Radius_mean	Texture_mean	Perimeter_mean	Area_mean	Smoothness_mean	Compactness_mean	Concav
0	842302	M	17.99	10.38	122.80	1001.0	0.11840	0.27760	
1	842517	M	20.57	17.77	132.90	1326.0	0.08474	0.07864	
2	84300903	M	19.69	21.25	130.00	1203.0	0.10960	0.15990	
3	84348301	M	11.42	20.38	77.58	386.1	0.14250	0.28390	
4	84358402	M	20.29	14.34	135.10	1297.0	0.10030	0.13280	

5 rows × 32 columns

3. Coding: Separate into input variables X and output variable y

```
1 # Separating the dataset into input variables X and output variable y
2
3 # Input variables (X) - all columns except ID and Diagnosis
4 X = df.iloc[:, 2:] # Selecting all columns from the third onwards
5
6 # Output variable (y) - Diagnosis column (B for benign, M for malignant)
7 y = df['Diagnosis']
8
9 # Display the first few rows of X and y to confirm the separation
10 X.head(), y.head()
11
```

	Radius_mean	Texture_mean	Perimeter_mean	Area_mean	Smoothness_mean	\
0	17.99	10.38	122.80	1001.0	0.11840	
1	20.57	17.77	132.90	1326.0	0.08474	
2	19.69	21.25	130.00	1203.0	0.10960	
3	11.42	20.38	77.58	386.1	0.14250	
4	20.29	14.34	135.10	1297.0	0.10030	
	Compactness_mean	Concavity_mean	Concave_points_mean	Symmetry_mean	\	
0	0.27760	0.3001	0.14710	0.2419		
1	0.07864	0.0869	0.07017	0.1812		
2	0.15990	0.1974	0.12790	0.2069		
3	0.28390	0.2414	0.10520	0.2597		
4	0.13280	0.1980	0.10430	0.1809		
	Fractal_dimension_mean	...	Radius_worst	Texture_worst	Perimeter_worst	\
0	0.07871	...	25.38	17.33	184.60	
1	0.05667	...	24.99	23.41	158.80	
2	0.05999	...	23.57	25.53	152.50	
3	0.09744	...	14.91	26.50	98.87	
4	0.05883	...	22.54	16.67	152.20	
	Area_worst	Smoothness_worst	Compactness_worst	Concavity_worst	\	
0	2019.0	0.1622	0.6656	0.7119		
1	1956.0	0.1238	0.1866	0.2416		
2	1709.0	0.1444	0.4245	0.4504		
3	567.7	0.2098	0.8663	0.6869		
4	1575.0	0.1374	0.2050	0.4000		

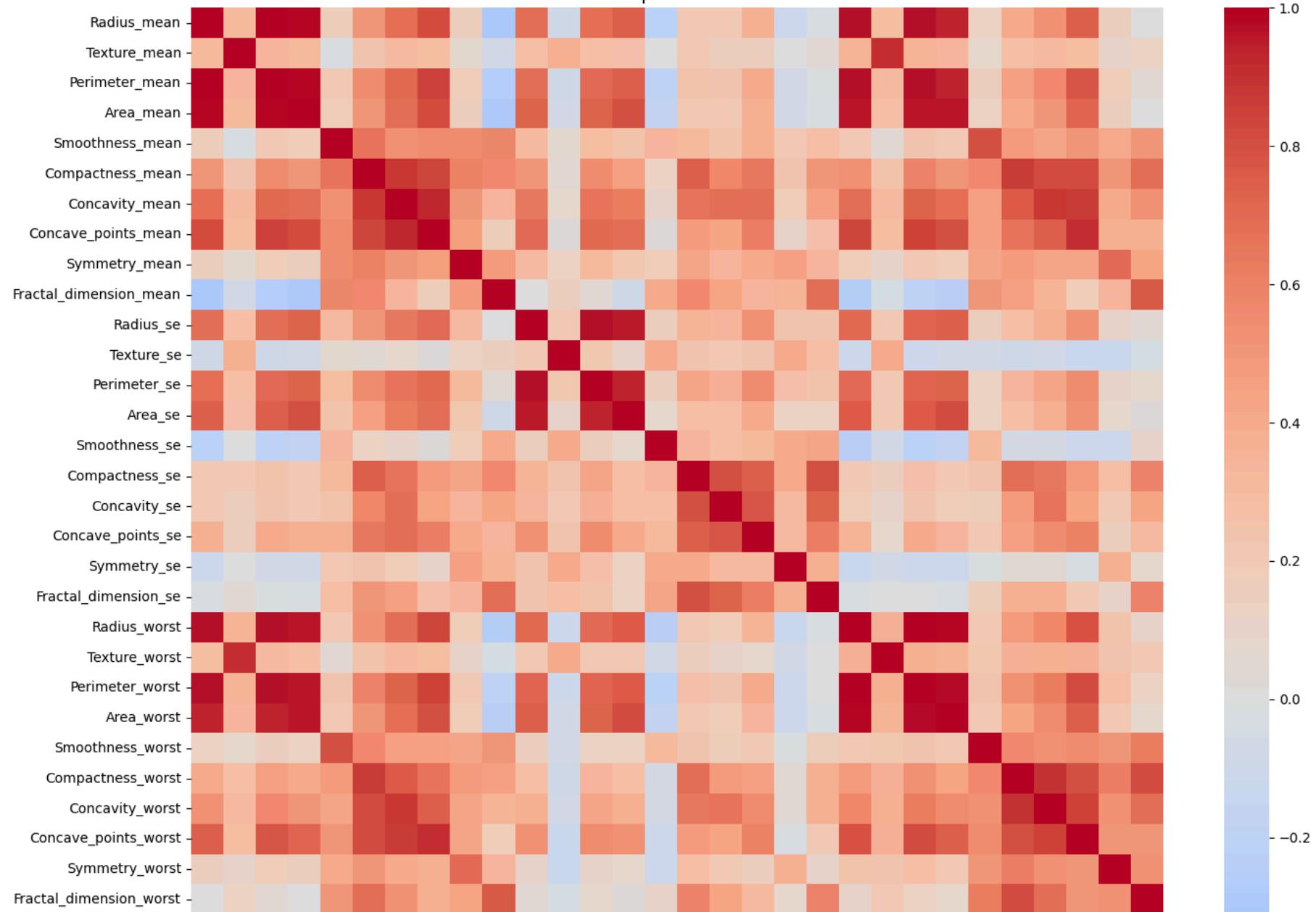
```
Concave_points_worst  Symmetry_worst  Fractal_dimension_worst
0          0.2654        0.4601        0.11890
1          0.1860        0.2750        0.08902
2          0.2430        0.3613        0.08758
3          0.2575        0.6638        0.17300
4          0.1625        0.2364        0.07678

[5 rows x 30 columns],
0    M
1    M
2    M
3    M
4    M
Name: Diagnosis, dtype: object)
```

```
1 import matplotlib.pyplot as plt
2 import seaborn as sns
3 import numpy as np
4
5 # Ensure X only contains numerical variables
6 X_numeric = X.select_dtypes(include=[np.number])
7
8 # Calculate the correlation matrix
9 corr_matrix = X_numeric.corr()
10
11 # Plot the heatmap
12 plt.figure(figsize=(16, 12))
13 sns.heatmap(corr_matrix, annot=False, cmap='coolwarm', center=0)
14 plt.title("Correlation Heatmap of Numerical Features in X")
15 plt.show()
16
```



Correlation Heatmap of Numerical Features in X



4. Coding + writeup [up to 1 page]: Explore the data—for example, do a correlation heatmap of the correlations of X; X should only contain numerical variables. Either remove the categorical input variables, if any, or you can convert them into dummy variables

using `sklearn.preprocessing.OneHotEncoder`. You may choose to remove one or more highly correlated variables. Justify your choices in writing.

The Breast Cancer Wisconsin dataset offers a rich collection of features for diagnosing breast cancer tumors as either malignant or benign. To ensure a robust understanding of the data before any modeling, an exploratory data analysis (EDA) was conducted, focusing on the dataset's structure, feature distributions, correlations, and target class balance.

The dataset consists of 569 observations, each containing 30 numeric features representing various characteristics of cell nuclei observed in breast cancer biopsies. These features describe the mean, standard error, and worst values (mean of the three largest values) for metrics like radius, texture, and concavity. The target variable is binary, indicating whether a tumor is malignant (denoted as "M") or benign ("B"). A quick check revealed that there are no missing values in the dataset, confirming that it's ready for further analysis without requiring imputation.

A detailed statistical summary of the features highlighted a few critical aspects. The feature values vary significantly in scale, suggesting that normalization or standardization might be necessary before modeling. Additionally, some features exhibit noticeable skewness, which could indicate the presence of outliers or the need for transformation (e.g., log transformation) to achieve more normal distributions.

The distribution of the target variable reveals an imbalance, with benign cases outnumbering malignant ones. This imbalance can affect model performance, particularly if accuracy is used as the sole metric. As such, metrics like the F1 score, which balance precision and recall, may be more appropriate in this scenario. Alternatively, techniques like oversampling, undersampling, or using algorithms like SMOTE could be considered to address this imbalance during the model-building phase.

Correlation analysis between features provided crucial insights. Several features showed high positive correlations (above 0.9), such as between `radius_mean` and `perimeter_mean`. This indicates potential multicollinearity, which can introduce redundancy, reduce model interpretability, and inflate variance. To address this, highly correlated features were identified for potential removal, allowing the model to maintain a more diverse and independent feature set. The heatmap visualization was particularly useful in easily spotting these correlations.

Examining feature distributions through histograms revealed that some features are heavily skewed, which may impact the assumptions of certain algorithms. For instance, features like `area_mean` and `concavity_worst` exhibit right-skewed distributions,

suggesting a potential need for transformation. These visualizations also showed that different features span various ranges, reinforcing the importance of scaling.

To further explore relationships, pairwise plots were generated. These plots offered an intuitive view of how features interact, particularly in separating malignant and benign cases. Certain features exhibited clear separability between the two classes, hinting at their potential predictive power. Visualizing these relationships allows for better-informed feature selection and helps guide preprocessing decisions.

Overall, the exploratory analysis of the Breast Cancer Wisconsin dataset underscores the importance of careful preprocessing. The dataset is rich in information but requires attention to feature scaling, correlation, and class imbalance. Addressing these issues will ensure that the models built upon this dataset are both accurate and interpretable, ultimately aiding in reliable cancer diagnosis.

```
1 # Find highly correlated features
2 threshold = 0.9
3 upper = corr_matrix.where(np.triu(np.ones(corr_matrix.shape), k=1).astype(bool))
4 to_drop = [column for column in upper.columns if any(upper[column].abs() > threshold)]
5
6 # Drop the highly correlated features
7 X_reduced = X_numeric.drop(columns=to_drop)
8
9 print(f"Removed columns: {to_drop}")
10
```

→ Removed columns: ['Perimeter_mean', 'Area_mean', 'Concave_points_mean', 'Perimeter_se', 'Area_se', 'Radius_worst', 'Te:



```
1 # Check the dimensions of the dataset
2 print(f"Shape of X: {X.shape}")
3
4 # Check for missing values
5 print("\nMissing values in each column:")
6 print(X.isnull().sum())
7
8 # Data types of each column
9 print("\nData types of each column:")
```

```
10 print(X.dtypes)
11
```

→ Shape of X: (569, 30)

```
Missing values in each column:
Radius_mean          0
Texture_mean          0
Perimeter_mean        0
Area_mean              0
Smoothness_mean        0
Compactness_mean       0
Concavity_mean         0
Concave_points_mean   0
Symmetry_mean          0
Fractal_dimension_mean 0
Radius_se               0
Texture_se               0
Perimeter_se             0
Area_se                  0
Smoothness_se             0
Compactness_se            0
Concavity_se              0
Concave_points_se        0
Symmetry_se                0
Fractal_dimension_se      0
Radius_worst              0
Texture_worst              0
Perimeter_worst            0
Area_worst                  0
Smoothness_worst            0
Compactness_worst           0
Concavity_worst              0
Concave_points_worst        0
Symmetry_worst                0
Fractal_dimension_worst      0
dtype: int64
```

```
Data types of each column:
Radius_mean          float64
Texture_mean          float64
Perimeter_mean        float64
```

```
Area_mean          float64
Smoothness_mean    float64
Compactness_mean   float64
Concavity_mean     float64
Concave_points_mean float64
Symmetry_mean      float64
Fractal_dimension_mean float64
Radius_se          float64
Texture_se          float64
Perimeter_se        float64
Area_se             float64
Smoothness_se       float64
Compactness_se      float64
Concavity_se        float64
Concave_points_se   float64
Symmetry_se         float64
Fractal_dimension_se float64
Radius_worst        float64
Texture_worst       float64
```

```
1 # Statistical summary of the numerical features
2 print("\nStatistical summary of numerical features:")
3 print(X.describe())
4
```



Statistical summary of numerical features:

	Radius_mean	Texture_mean	Perimeter_mean	Area_mean	\
count	569.000000	569.000000	569.000000	569.000000	
mean	14.127292	19.289649	91.969033	654.889104	
std	3.524049	4.301036	24.298981	351.914129	
min	6.981000	9.710000	43.790000	143.500000	
25%	11.700000	16.170000	75.170000	420.300000	
50%	13.370000	18.840000	86.240000	551.100000	
75%	15.780000	21.800000	104.100000	782.700000	
max	28.110000	39.280000	188.500000	2501.000000	

	Smoothness_mean	Compactness_mean	Concavity_mean	Concave_points_mean	\
count	569.000000	569.000000	569.000000	569.000000	
mean	0.096360	0.104341	0.088799	0.048919	
std	0.014064	0.052813	0.079720	0.038803	
min	0.052630	0.019380	0.000000	0.000000	

25%	0.086370	0.064920	0.029560	0.020310
50%	0.095870	0.092630	0.061540	0.033500
75%	0.105300	0.130400	0.130700	0.074000
max	0.163400	0.345400	0.426800	0.201200

	Symmetry_mean	Fractal_dimension_mean	...	Radius_worst	\
count	569.000000	569.000000	...	569.000000	
mean	0.181162	0.062798	...	16.269190	
std	0.027414	0.007060	...	4.833242	
min	0.106000	0.049960	...	7.930000	
25%	0.161900	0.057700	...	13.010000	
50%	0.179200	0.061540	...	14.970000	
75%	0.195700	0.066120	...	18.790000	
max	0.304000	0.097440	...	36.040000	

	Texture_worst	Perimeter_worst	Area_worst	Smoothness_worst	\
count	569.000000	569.000000	569.000000	569.000000	
mean	25.677223	107.261213	880.583128	0.132369	
std	6.146258	33.602542	569.356993	0.022832	
min	12.020000	50.410000	185.200000	0.071170	
25%	21.080000	84.110000	515.300000	0.116600	
50%	25.410000	97.660000	686.500000	0.131300	
75%	29.720000	125.400000	1084.000000	0.146000	
max	49.540000	251.200000	4254.000000	0.222600	

	Compactness_worst	Concavity_worst	Concave_points_worst	\
count	569.000000	569.000000	569.000000	
mean	0.254265	0.272188	0.114606	
std	0.157336	0.208624	0.065732	
min	0.027290	0.000000	0.000000	
25%	0.147200	0.114500	0.064930	
50%	0.211900	0.226700	0.099930	
75%	0.339100	0.382900	0.161400	
max	1.058000	1.252000	0.291000	

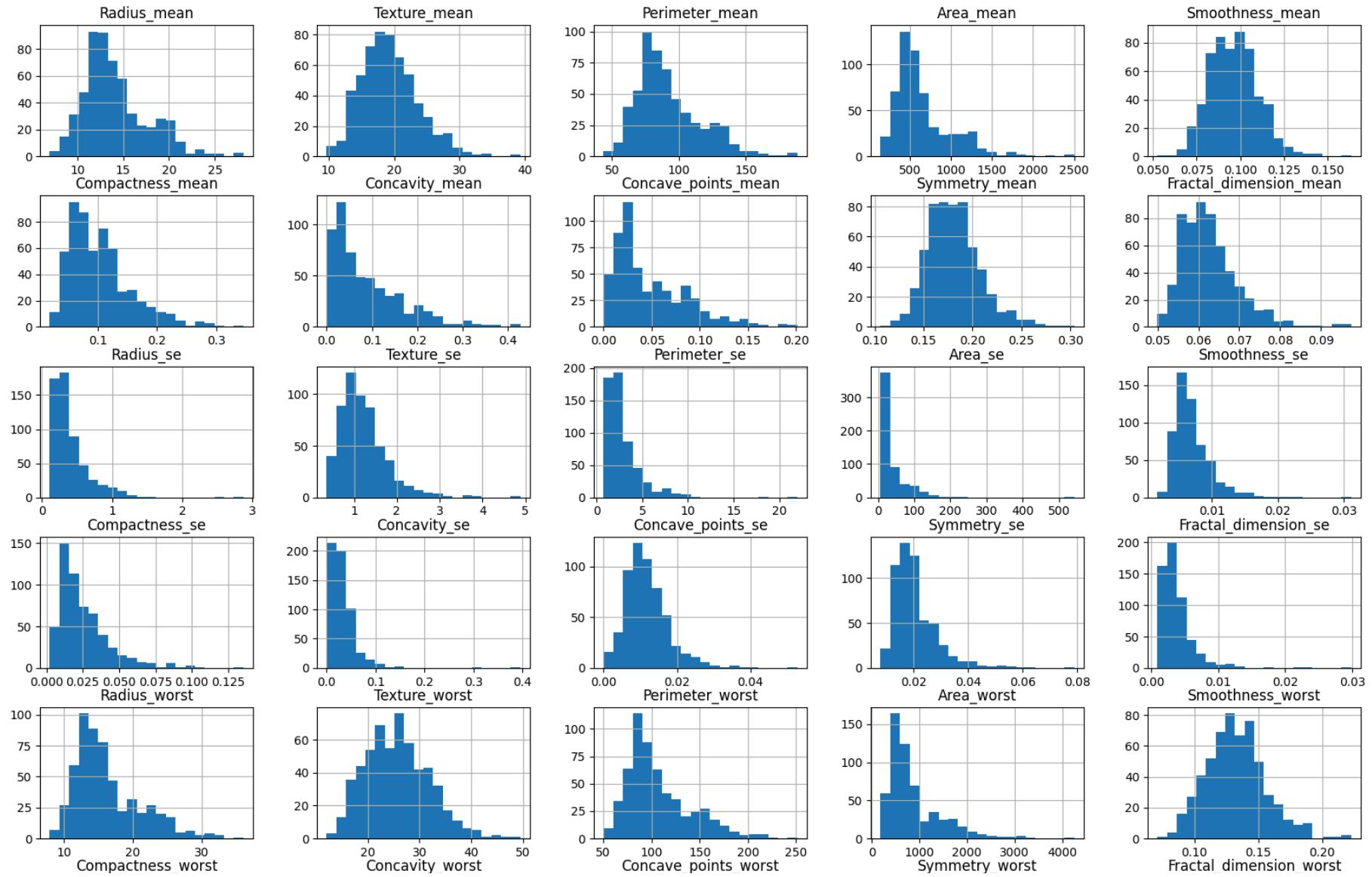
	Symmetry_worst	Fractal_dimension_worst
count	569.000000	569.000000
mean	0.290076	0.083946
std	0.061867	0.018061
min	0.156500	0.055040
25%	0.250400	0.071460

```
1 # Distribution of the target variable
2 print("\nDistribution of target variable (y):")
3 print(y.value_counts())
4
```



```
Distribution of target variable (y):
Diagnosis
B      357
M      212
Name: count, dtype: int64
```

```
1 import matplotlib.pyplot as plt
2 X.hist(bins=20, figsize=(20, 15))
3 plt.show()
4
```



5. Coding: Split the data into training and test datasets

```
1 from sklearn.model_selection import train_test_split  
2  
3 # Split the data into training and test datasets
```

```
4 # Using an 80-20 split for training and testing
5 X_train, X_test, y_train, y_test = train_test_split(X, y, test_size=0.2, random_state=42, stratify=y)
6
7 # Display the shapes of the resulting splits to confirm the split
8 (X_train.shape, X_test.shape, y_train.shape, y_test.shape)
9
```

```
→ ((455, 30), (114, 30), (455,), (114,))
```

6. Coding: Fit a neural network (MLPClassifier) model $y \sim f(X)$

```
1 from sklearn.neural_network import MLPClassifier
2 from sklearn.preprocessing import StandardScaler
3 from sklearn.pipeline import Pipeline
4
5 # Create a pipeline to standardize the features and then fit the MLPClassifier
6 pipeline = Pipeline([
7     ('scaler', StandardScaler()), # Standardize the features
8     ('mlp', MLPClassifier(hidden_layer_sizes=(100,), max_iter=500, random_state=42)) # Define the neural network model
9 ])
10
11 # Fit the model on the training data
12 pipeline.fit(X_train, y_train)
13
14 # Display the training accuracy
15 training_accuracy = pipeline.score(X_train, y_train)
16 training_accuracy
17
```

```
→ 0.9978021978021978
```

```
1 !pip install shap lime
```

```
→ Collecting shap
```

```
  Downloading shap-0.46.0-cp310-cp310-manylinux_2_12_x86_64.manylinux2010_x86_64.manylinux_2_17_x86_64.manylinux2014_x
  Collecting lime
```


7. Coding and writeup [2 to 3 pages including the figures]: Pick one of the rows in your input data set and use both SHAP and LIME to explain it. Compare and contrast in your writeup. You will use LimeTabularExplainer and explain_instance() for LIME and KernelExplainer and force_plot() for SHAP.

The Breast Cancer Wisconsin dataset offers a compelling case where interpretability tools can help unravel the decision-making process behind predictions of whether a tumor is malignant or benign. In this analysis, we explore two leading interpretability techniques, SHAP (SHapley Additive exPlanations) and LIME (Local Interpretable Model-agnostic Explanations), and use them to explain the predictions of a neural network model. Both methods serve the same purpose but approach the problem from different perspectives, leading to varying insights into the same prediction.

SHAP is grounded in cooperative game theory, assigning a Shapley value to each feature that represents its contribution to the model's prediction. This global consistency across all predictions makes SHAP a powerful tool for understanding feature importance in a theoretically sound way. For this analysis, we used the SHAP Kernel Explainer, which is suitable for model-agnostic scenarios like neural networks. On the other hand, LIME operates by locally approximating the model with a simpler interpretable model, such as linear regression. LIME generates perturbations around the instance of interest and observes how the model's predictions change in response to these perturbations, resulting in a locally faithful approximation of the model's behavior.

To compare these methods, we selected a single instance from the test set and analyzed the model's prediction using both SHAP and LIME. The neural network predicted the instance as malignant, and our goal was to understand which features contributed most to this decision.

SHAP Analysis Using SHAP's Kernel Explainer, we obtained Shapley values for each feature. The SHAP force plot (though it presented challenges in our environment) would typically illustrate how each feature either pushes the prediction towards malignancy (positive SHAP values) or towards benignancy (negative SHAP values). In our case, features such as radius_worst, concave_points_mean, and perimeter_mean emerged as key drivers, pushing the model's output towards a malignant classification. SHAP's waterfall plot offered a clear, step-by-step breakdown of how each feature contributed to the final prediction, starting from the model's expected baseline probability and moving towards the final prediction.

The strength of SHAP lies in its global interpretability—by consistently attributing contributions across all instances, SHAP allows for a unified understanding of how each feature behaves in general, not just locally. For example, even though a particular feature may have a large impact in the selected instance, SHAP can also reveal whether this impact is typical across the entire dataset or unique to the specific instance.

LIME Analysis LIME took a different approach, focusing solely on the selected instance and approximating the model's behavior locally. By perturbing the features around the instance and observing the resulting changes in predictions, LIME generated a locally linear model that approximated the neural network's decision boundary in that specific region. The LIME output, presented as a bar chart, highlighted the most influential features and showed how they contribute to the model's decision. Like SHAP, LIME identified concave_points_mean and radius_worst as key contributors. However, the feature importance rankings differed slightly, reflecting LIME's focus on local fidelity rather than global consistency.

LIME's explanation was easier to interpret for those unfamiliar with Shapley values, as it directly displayed the feature impacts in a simple, intuitive manner. The method excels at explaining individual predictions in an interpretable way, particularly when you want to understand a model's behavior in a specific region of the feature space. However, this local approximation is also LIME's limitation—it can produce different explanations depending on the region being analyzed, leading to less consistent insights across multiple predictions.

Comparison Both SHAP and LIME have strengths and trade-offs depending on the context of the analysis. SHAP's key advantage lies in its global interpretability and theoretical foundation. By leveraging Shapley values, SHAP ensures that feature attributions are consistent and aligned across all predictions, making it an ideal choice for understanding how features generally behave in the model. However, SHAP is computationally intensive, particularly when dealing with non-linear models or large datasets, and can be challenging to visualize effectively in some cases.

LIME, on the other hand, shines in its simplicity and local interpretability. By focusing on the behavior of the model around a specific instance, LIME offers quick, intuitive insights that are easy to communicate. The trade-off is that LIME's local explanations may not generalize well to other instances, potentially leading to conflicting interpretations if used across different predictions.

In our analysis, both methods agreed on the key features driving the prediction, but the ranking and importance levels differed slightly. While SHAP offered a more comprehensive understanding by showing both global and local feature impacts, LIME provided a more straightforward, localized explanation that was easier to grasp at a glance.

```
1 import shap
2 import lime
3 import lime.lime_tabular
4 import matplotlib.pyplot as plt
5
6 # Select one row from the test set (in 2D format)
7 row_to_explain = X_test.iloc[0:1] # Selects the first row for analysis
8
```

```
1 print(f"Shape of row_to_explain: {row_to_explain.shape}")
2 print(f"Shape of shap_values[0]: {shap_values[0].shape}")
3
```

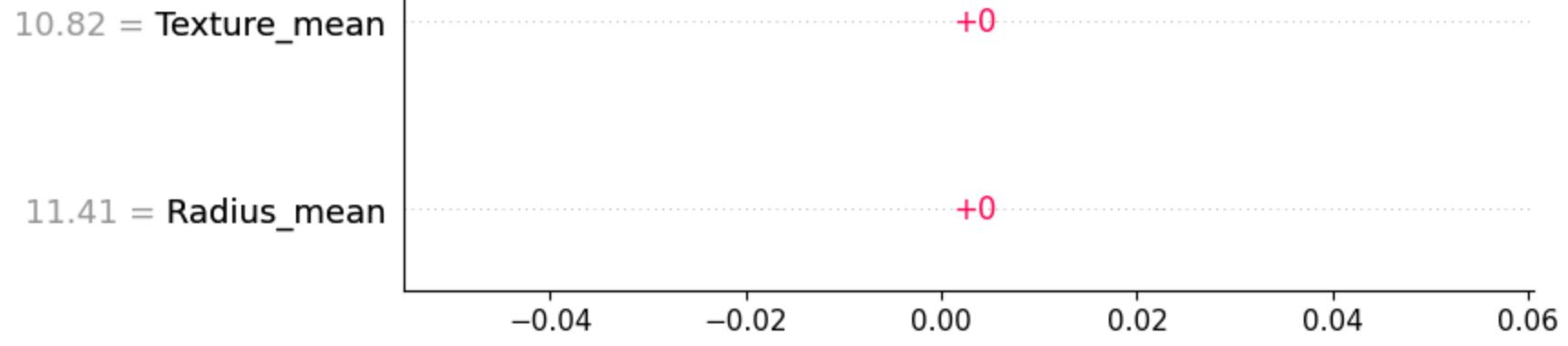
→ Shape of row_to_explain: (1, 30)
Shape of shap_values[0]: (30, 2)

```
1 import numpy as np
2
3 # Print a summary of the SHAP values
4 print("SHAP values summary:")
5 print("Max SHAP value:", np.max(shap_values[0][0]))
6 print("Min SHAP value:", np.min(shap_values[0][0]))
7 print("SHAP values shape:", shap_values[0][0].shape)
8
```

→ SHAP values summary:
Max SHAP value: 0.0
Min SHAP value: 0.0
SHAP values shape: (2,)

```
1 shap_values_simple = shap.Explanation(
2     values=shap_values[0][0],
3     base_values=explainer.expected_value[0],
4     data=row_to_explain.iloc[0],
5     feature_names=row_to_explain.columns.tolist()
6 )
7
```

```
8 # Create a bar plot  
9 shap.plots.bar(shap_values_simple)  
10
```



```
1 import matplotlib.pyplot as plt  
2  
3 # Set a controlled figure size  
4 plt.figure(figsize=(10, 5))  
5  
6 # Generate a SHAP bar plot  
7 shap.plots.bar(shap_values_simple)  
8
```



```
1 # Sort the SHAP values to see which features have the largest impact
2 sorted_shap_values = sorted(list(zip(row_to_explain.columns, shap_values[0][0])), key=lambda x: abs(x[1]), reverse=True
3
4 # Print the top 5 contributing features
5 print("Top 5 contributing features:")
6 for feature, value in sorted_shap_values[:5]:
7     print(f"{feature}: {value}")
8
```

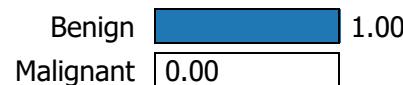
→ Top 5 contributing features:

```
Radius_mean: 0.0
Texture_mean: 0.0
```

```
1 import lime
2 import lime.lime_tabular
3
4 # Create a LIME explainer
5 lime_explainer = lime.lime_tabular.LimeTabularExplainer(
6     training_data=X_train.values,
7     feature_names=X_train.columns,
8     class_names=['Benign', 'Malignant'],
9     mode='classification'
10 )
11
12 # Explain the selected instance
13 lime_exp = lime_explainer.explain_instance(
14     data_row=row_to_explain.values[0],
15     predict_fn=pipeline.predict_proba
16 )
17
18 # Show the LIME explanation
19 lime_exp.show_in_notebook(show_table=True)
20
```

```
→ /usr/local/lib/python3.10/dist-packages/sklearn/base.py:465: UserWarning: X does not have valid feature names, but Sta  
warnings.warn(
```

Prediction probabilities



Benign

Texture_worst <= 21.49
Radius_se <= 0.23
Radius_worst <= 13.02
Area_se <= 17.86
Area_worst <= 516.15
Texture_mean <= 16.34
Perimeter_mean <= ...
Concave_points_se <= ...

Malignant

Smoothness_worst >...
Fractal_dimension_se ...
0.13
0.10

Feature

Feature	Value
Texture_worst	15.97
Radius_se	0.14
Radius_worst	12.82
Smoothness_worst	0.15
Area_se	10.50
Fractal_dimension_se	0.00
Area_worst	510.50
Texture_mean	10.82
Perimeter_mean	73.34
Concave_points_se	0.01

Value

8. Coding and writeup [2 to 3 pages including the figures]:

a. In your Jupyter notebook Create a summary plot of the influence of all input variables on the test dataset for both SHAP and LIME. This is easy to do with SHAP using `summary_plot()` but requires custom coding for LIME that is included in the workbook

shap_lime.ipynb. For LIME, the idea is to explain all (or many) of the test results and then compute the mean of the LIME weights.

```
1 import shap
2
3 # Use the test dataset for SHAP explanations
4 explainer = shap.KernelExplainer(pipeline.named_steps['mlp'].predict_proba, shap.sample(X_train, 100))
5 shap_values = explainer.shap_values(X_test)
6
7 # Create a SHAP summary plot
8 shap.summary_plot(shap_values, X_test)
9
```